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(71) Applicant: STICHTING VRIENDEN VAN DE STICHTING
DR. KARL LANDSTEINER
Plesmanlaan 125
NL-1066 CX Amsterdam(NL)

(72) Inventor: Pannekoek, Hans
Stommeerweg 36
NL-1431 EW Aalsmeer(NL)

(72) Inventor: Verwey, Cornelis Lammert
Stationsstraat 20
NL-1391 GN Abcoude(NL)

(72) Inventor: Diergaarde, Paul Johan
Rode Kruislaan 1005
NL-1111 ZX Diemen(NL)

(72) Inventor: Hart, Margaretha Hendrika L.
De Schaapherderstraat 34 (III)
NL-1061 GW Amsterdam(NL)

(74) Representative: van der Beek, George Frans et al.
Nederlandsch Octrooibureau Johan de Wittlaan 15 P.O.
Box 29720
NL-2502 LS 's-Gravenhage(NL)

(54) Preparation of the human von Willebrand factor by recombinant DNA.

(57) Full length human von Willebrand factor (vWF) cDNA was assembled from partial, overlapping vWF cDNAs. This cDNA construct includes an "open" reading frame of 2,813 amino-acid residues, apparently representing a (pre)-pro-sequence of 763 and mature vWF of 2,050 amino acids. Both the pro-sequence and mature vWF harbor an arg-gly-aspartic ("R-G-D") sequence which has been implicated in cell attachment functions. It is assumed that the pro-sequence is equivalent to von Willebrand Antigen II (vW AgII). The (glyco)-proteins prepared by cultivation of microorganisms or other hosts like animal or human cells containing a plasmide or phage with said cDNA-coding may be used in pharmaceutical compositions intended for administration to patients with the von Willebrand's disease.

cDNA coding for the human von Willebrand factor, plasmids or phages, preparation of proteins by the cultivation of the above mentioned hosts, the obtained proteins and pharmaceutical compositions containing a biologically active form of the obtained proteins.

10 The von Willebrand factor (vWF) is a large, multimeric plasma protein, composed of an apparently, single glycoprotein with a molecular weight (MW) of about 225 kD. These subunits are linked together by disulfide bonds. In plasma vWF circulates as multimers, ranging from dimers to multimers of more than 50 subunits. Dimers
15 consist of two subunits joined, probably at their C-termini, by flexible "rod-shaped" domains and are presumed to be the protomers in multimerization. The protomers are linked through large, probably N-terminal, globular domains to form multimers.

vWF is synthesized by endothelial cells and megakaryocytes. It
20 is believed that this protein is initially produced as a 260 kD glycosylated precursor that is subsequently subjected to carbohydrate processing, sulfatation, dimerization, multimerization and to proteolytic cleavage to yield the mature 225 kD subunit (Sporn, L.A., et al. (1985) Biosynthesis of vWF protein in human megakaryocytes, J. Clin. Invest. 76, 1102-1106); Wagner, D.D., et al. (1984)
25 J. of Cell Biology 99, 2123-2130). It has been shown that vWF is stored in the Weibel-Palade bodies within the endothelial cells. It can not be excluded that these organelles play a role in the processing of the precursor protein.

30 vWF participates in critical steps in hemostasis. It is involved in platelet-vessel wall interactions after vascular injury, leading to platelet plug formation. On the vWF protein domains have been assigned which show specific interaction with the platelet glycoproteins IB (Jenkins, C.S.P., et al. (1976) J. Clin. Invest. 69, 1212-1222), IIB/IIIA (Fujimoto, T., et al. (1982) Adenine
35 diphosphate induces binding of von Willebrand factor to human platelets, Nature 297, 154-156), collagens type I and III and with

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another, yet unidentified, component in the subendothelium. These assignments are based on studies with monoclonal anti-vWF antibodies which are able to inhibit a particular interaction of vWF. For a in depth analysis of structure-function relationships of the vWF protein, a full length vWF cDNA will be indispensable. Introduction of well-defined mutations within this cDNA and expression of the mutated cDNA in a suitable host will allow a detailed localization of functional domains within the vWF protein. Recently, applicants and others (Lynch, D.C., et al. (1985) Cell 41, 49-56; Ginsburg, M., et al. (1985) J. Biol. Chem. 260, 3931-3936; Verweij, C.L., et al. (1985) Nucleic Acids Research 13, 4699-4717, based on Dutch patent application 85.00961; Sadler, J.E., et al. (1985) Proc. Natl. Acad. Sci. USA 82, 6394-6398) have cloned partial vWF cDNA sequences. The presence of a short 3' untranslated region (136 nt) on vWF mRNA, which extends to about 9,000 nt, led us to assume that the precursor protein for vWF has a MW considerable larger than the reported 260 kD (Wagner, D.D., et al. (1984) J. of Cell Biology 99, 2123-2130). A full length vWF cDNA will enable us to elucidate the enigma on the MW of the precursor, characterize its processing pathway and establish the primary structure.

This invention relates to the isolation and the nucleotide sequence of cDNAs, spanning the entire vWF mRNA, and on the assembly of these sequences into a full length, functional vWF cDNA. Further this invention relates to plasmides and phages containing above indicated vWF cDNAs as well as to the microorganisms like bacteria and fungi as well as animal or human cells containing said plasmids or phages. Moreover the proteins prepared by cultivating above indicated hosts and the pharmaceutical compositions containing a biologically active form of the obtained proteins do also fall within the scope of the present invention. In this respect it is emphasized that by means of such pharmaceutical compositions it is possible to circumvent the safety problems bound to the administration of vWF protein recovered from blood samples of normal individuals.

To perform the investigations forming the basis of the invention, use has been made of the materials and method below.

MATERIALS AND METHODS

cDNA cloning:

Total RNA was purified from in vitro cultured endothelial cells, derivated from veins of human umbilical cords (Verweij, C.L., et al. (1985) Nucleic Acids Research 13, 4699-4717, based on Dutch patent application 85.00961). Primer-directed cDNA was synthesized essentially according to a protocol described (Gubler, U., et al. (1983) Gene 25, 263-269; Toole, J.J., et al. (1985) Nature 312, 342- 347). The cDNA synthesis was arrested by adding EDTA and SDS till a final concentration of, respectively, 20 mM and 0.1 %. The cDNA preparations were extracted with phenol-chloroform, then precipitated with ethanol and purified by chromatography on Sephadex G-50. In the case of primer-directed cDNA synthesis with primer A (5' CACAGGCCACACGTGGGAGC 3'), complementary to nucleotides 6901 till 6921, the cDNA preparation was digested with BglIII (positions 6836 and 2141) and KpI (position 4748). Subsequently, the digested cDNA was size-fractionated by chromatography on a Sepharose CL-4B column. Fractions containing cDNA larger than about 600 bp were ligated to plasmid pMBL11, digested with BglIII and KpnI. A cDNA library of about 15,000 independent colonies was established, using strain E.coli DH1 as a host, which was screened with two oligonucleotide probes (B and C). Probe B (5' GAGGCAGGATTTCCGGTGAC 3'), complementary to nucleotides 4819 till 4839 was employed for the isolation of the plasmid pvWF2084, harboring a 2,084 bp BglIII-KpnI vWF cDNA fragment, while probe C (5' CAGGGACACCTTTCAGGGC 3'), complementary to 2467 till 2487, was used for the detection of plasmid pvWF2600, harboring an approximately 2,600 bp KpnI-BglIII vWF cDNA fragment.

Using probe C for primer-directed synthesis applicants divided the resulting cDNA preparation in two parts. One part was C-tailed and annealed to G-tailed plasmid pUC9 as described before (Verweij, C.L., et al. (1985) Nucleic Acids Research 13, 4699-4717, based on Dutch patent application 85.00961) and used to transform E.coli DH1. Six thousand independent colonies were hybridized with a "nick translated" 575 bp BglIII-BamHI vWF cDNA fragment of pvWF2600 DNA. A positive clone, harboring a plasmid with the longest insert (about 1,800 bp, designated pvWF1800) was chosen for further study. The

ther part of the primer 3-directed cDNA preparation was treated with EcoRI methylase and subsequently with T4-DNA polymerase and dNTPs to ensure blunt-ended termini (Maniatis, T., et al. (1982) Molecular Cloning Laboratory Manual, Cold Spring Harbor Laboratory). Phosphorylated EcoRI linkers (New England Biolabs, Beverly, MA) were ligated to the termini of the cDNA preparation and unreacted components were removed by Sephadex G-50 chromatography. The XhoI site, located about 350 bp downstream of the 5' end of the vWF cDNA insert of pvWF1800 DNA, was used for another selection. After digestion with an excess of EcoRI and XhoI chromatography on Sepharose CL-4B was employed to remove digested EcoRI linkers. The final preparation was ligated to plasmid pMBL11 DNA which had been digested with EcoRI plus XhoI and used to transform E.coli DH1. A collection of about 10,000 independent colonies was hybridized with a "nick translated" 350 bp XhoI-HindIII vWF cDNA fragment from plasmid pvWF1800. The HindIII site of this fragment has been derived from the polylinker of the vector pUC9. A positive clone, harboring the longest insert (about 1,330 bp, designated pvWF1330) was further studied.

S1 nuclease protection analysis:

Applicants used a probe for S1 nuclease protection experiments an XhoI-EcoRI fragment of about 5,300 bp (probe V) from plasmid pvWF2600 which contains a 2,390 bp segment (XhoI-KpnI) constituted of vWF cDNA (Fragment V, Figure 1). Probe II was a 4,800 bp XbaI-EcoRI fragment from plasmid pvWF1330 which harbors a 735 bp XbaI-XhoI vWF cDNA segment (Fragment II, Figure 1). The fragments were 3' end labelled, using DNA polymerase I (large fragment) (New England Biolabs, Beverly, MA) to fill in recessed ends (Maniatis, T., et al. (1982) Molecular Cloning Laboratory Manual, Cold Spring Harbor Laboratory). Subsequently, these probes were isolated by electrophoresis on a 0.7 % low-melting agarose gel and purified as described (Wieslander, L. (1979) Anal. Biochem. 48, 305-309).

Three other vWF cDNA fragments were subcloned in double stranded M13mpl8 and employed as probes. To that end the anti-sense

pvWF1500 (Fragment III, Figure 1), a 385 bp XbaI-EcoRI fragment of plasmid pvWF1330 (Fragment I, Figure 1) and a 575 bp BamHI-BglII fragment of plasmid pvWF2600 (Fragment IV, Figure 1). After DNA synthesis, initiated at the M13 primer, double stranded DNA was digested with both HindIII and PvuII for fragment III (to yield probe II), with both XbaI and EcoRI for fragment I (to yield probe I) and with both BamHI and PvuII for fragment IV (to yield probe IV). The rationale for the construction of probes II, III, IV and V is that they contain a segment of vector DNA non-complementary with endothelial RNA. For example probes III and IV harbor about 200 bp, derived from M13mpl8. These probes were subjected to electrophoresis on a 5 % polyacrylamide - 8M urea gel and the fragments of interest were isolated (Maxam A.G., et al. (1980) Methods Enzymol. 65, 499-560).

S1 nuclease protection experiments were carried out essentially as described (Berk, A.J., et al. (1977) Cell 12, 721-732). One microgram of human endothelial polyA RNA was added to 10,000 - 100,000 counts per min. of radiolabelled probe, heated for 10 min. at 80°C, followed by an incubation overnight at 60°C for probes I, II, III and V and at 57°C for probe IV. Digestion with 200 Units of S1 nuclease (Bethesda Research Lab., Gaithersburg, Md.) per ml for 20 min. at 45°C. Undigested DNA was precipitated with ethanol and the pellets were dissolved in the appropriate loading buffer for electrophoresis on a 0.8 % alkaline agarose gel or on a 6 % polyacrylamide sequencing gel (Maniatis, T., et al. (1982) Molecular Cloning Laboratory Manual, Cold Spring Harbor Laboratory). The first procedure was employed for probes I and III, whereas the second one was applied for probes II, IV and V.

Assembly of full length vWF cDNA:

For the construction of plasmid pSP6330vWF, harboring a continuous vWF cDNA segment of about 6,331 bp extending from the HindIII site (position 2235) till the SacI site within the 3' untranslated region (position 8562) the following vWF cDNA fragments were isolated: the 2,517 bp HindIII-KpnI fragment (position 2236 till 4753) from pvWF2600 DNA, the 2,084 bp KpnI-BglII fragment (position 4753 till 6837) from pvWF2084 DNA and the 1,730 bp BglII-SacI fragment (position 6837 till 8567) from pvWF2280 DNA. These

three vWF cDNA fragments were ligated simultaneously into the vector pSP64 (Melton, D.A., et al. (1984) Nucl. Acids Research 12, 7035-7056), digested with both HindIII and SacI. About half of the resulting transformants contained a plasmid (denoted pSP6330vWF) with the desired vWF cDNA insert of about 6,331 bp, as verified by restriction enzyme analysis.

For the construction of plasmid pSP8800vWF, harboring full length vWF cDNA, the following fragments were isolated: the 6,330 bp HindIII-EcoRI insert of plasmid pSP6330vWF (the EcoRI site is derived from the polylinker present on the vector), the 1,044 bp XhoI-HindIII fragment (position 1092 till 2236) from pWFl800 and the 1,327 bp EcoRI-XhoI fragment (position - 236 till 1092) from pWFl330. A five-fold molar excess of each of these three fragments were again ligated simultaneously with vector pSP65 DNA, cleaved with EcoRI and treated with calf intestine alkaline phosphatase (Boehringer, Mannheim, BRD). About 30 % of the resulting colonies harbored a plasmid with the desired, full length vWF cDNA insert of about 8,795 bp, as verified by restriction enzyme analysis and nucleotide sequence analysis.

In vitro transcription and translation:

In vitro transcription of linear SP6-based DNA templates with SP6 RNA polymerase (New England Nuclear, Dreieich, BRD) was performed in the presence of 0.1 mM UTP, CTP and ATP, 0.05 mM GTP and 2 mM of m7G(5')ppp(5')G (Pharmacia, Uppsala, Sweden) to provide mRNA preparations with a capped terminus (Melton, D.A., et al. (1984) Nucl. Acids Research 12, 7035-7056). In vitro translation of such 5' capped mRNAs was done in a rabbit reticulocyte lysate system (New England Nuclear, Dreieich, BRD), according to the manufacturer's specifications.

Analysis of the in vitro translation products was performed by electrophoresis on a 8 % SDS-polyacrylamide gel as described (Laemmli, U.K. (1970) Nature 227, 650-655).

RESULTS

Construction of partial vWF cDNA clones and assembly of full length

struction of plasmids containing part of a full length human vWF cDNA (Verweij, G.L., et al. (1985) *Nucleic Acids Research* 13, 4699-4717, based on Dutch patent application 85.00961; Lynch, D.C., et al. (1985) *Cell* 41, 49-56; Ginsburg, M., et al. (1985) *J. Biol. Chem.* 260, 3931-3936; Sadler, J.E., et al. (1985) *Proc. Nucl. Acad. Sci. USA* 82, 6394-6398). The most extended vWF cDNA, that applicants obtained from an oligo(dT)-primed human endothelial cDNA library, comprised about 2,280 bp (pvWF2280). Nucleotide sequence analysis revealed that this cDNA insert has been initiated at the polyA tail of vWF mRNA. To construct a full length vWF cDNA, applicants have isolated additional, overlapping vWF cDNA sequences which are located upstream of pvWF2280. For that purpose two biochemical selections were employed to enrich for the number of vWF cDNA harboring plasmids. First, oligonucleotide primers, derived from the partial nucleotide sequence (Sadler, J.E., et al. (1985) *Proc. Nucl. Acad. Sci. USA* 82, 6394-6398), were synthesized to direct cDNA synthesis with human endothelial polyA RNA as substrate. Second, cDNA preparations were digested with particular restriction endonucleases, known to dissect vWF cDNA at a limited number of sites (Ginsburg, M., et al. (1985) *J. Biol. Chem.* 260, 3931-3936; Sadler, J.E., et al. (1985) *Proc. Nucl. Acad. Sci. USA* 82, 6394-6398). These restriction sites can subsequently be employed to assemble a full length vWF cDNA. The cloning strategy is outlined in Figure 1A. The plasmids, containing adjacent vWF cDNA sequences, were designated pvWF1330, pvWF1800, pvWF2600, pvWF2084 and pvWF2280. The nucleotide sequence of the 5' part of pvWF1330 DNA (corresponding with the 5' end of vWF mRNA) revealed that nonsense codons were present in all three reading frames. From this finding applicants conclude that pvWF1330 DNA extends beyond the translation initiation codon.

S1 nuclease protection experiments with human endothelial RNA were performed to proof that the various vWF cDNA inserts are fully complementary to vWF mRNA. The construction of the probes and the conditions used are described in the section Materials and methods. The results are shown in Figure 2. In all cases the length of the protected fragments is in accord with the predicted length of the different probes. From these data applicants conclude that the vWF

5 cDNA inserts of, respectively, pWVF1330, pWVF1300 and pWVF2600 DNA are entirely complementary with vWF mRNA. The nucleotide sequence of the remaining cDNA inserts of plasmids pWVF2084 and pWVF2280 were shown before to correspond with the published sequence (Sadler, J.E., et al. (1985) Proc. Natl. Acad. Sci. USA 82, 6394-6398). Consequently, the different, adjacent vWF cDNA sequences are genuine copies of vWF mRNA.

10 The vWF cDNA sequences that applicants have constructed span a length of about 8,900 bp. This length is consistent with the size of vWF mRNA, determined by Northern blot analysis of human endothelial (polyA) RNA (Verweij, C.L., et al. (1985) Nucleic Acids Research 13, 4699-4717, based on Dutch patent application 85.00961). A detailed description of the assembly of full length vWF cDNA is given in the section Materials and methods. The correct composition of the assembled, full length vWF cDNA was established by restriction enzyme analysis.

15 Nucleotide sequence of full length vWF cDNA:

20 Nucleotide sequence analysis of vWF cDNA fragments was carried out both by the chemical degradation method (Maxam, A.M. et al., 1977) and by the dideoxy chain-termination procedure (Sanger, F., et al. (1977) Proc. Natl. Acad. Sci. USA 74, 5463-5467), according to the scheme outlined in Figure 1A. In Figure 3 the nucleotide sequence of about 8806 residues, extending from the 5' end of vWF mRNA, and the corresponding predicted amino acid sequence are presented. The remaining nucleotide sequence of the 3' part of vWF mRNA has been reported before (Sadler, J.E. et al., (1985) Proc. Natl. Acad. Sci. USA 82, 6394-6398; Verweij, C.L. et al., (1985) Nucleic Acids Research 13, 4699-4717, based on Dutch patent application 85.00961). In general the overlapping part of our nucleotide sequence of vWF cDNA with that of Sadler, J.E. et al., (1985) Proc. Natl. Acad. Sci. USA 82, 6394-6398 reveals no differences. However, the first 12 nucleotides (corresponding with position 2217 till 2229) at the 5' terminus of vWF cDNA on phage

independent, sequencing. No insert or deletion was observed at position 2309. Our analysis reveals a C residue,

whereas Sadler, J.E. et al., (1985) Proc. Natl. Acad. Sci. USA 82, 6394-6398 report an A residue, resulting in, respectively, a proline and a histidine at that particular position. This difference might be due to a polymorphism in the vWF gene, although the proline residue has been established independently by automated amino acid sequence analysis of the mature vWF protein (Hessel, E. et al., (1984) Thrombosis Research 35, 637-651).

The total length spanned by the adjacent vWF cDNA sequences is 8,814 bp, excluding the polyA tail of vWF mRNA. The translation initiation site was assigned to the ATG codon indicated (position 1 till 4), being the first initiator codon downstream of the TAG nonsense codon (position - 79 till - 82), which is followed by an "open" translation reading frame of 8,439 nucleotides (deduced from our sequence data and those of Sadler, J.E. et al., (1985) Proc. Natl. Acad. Sci. USA 82, 6394-6398). This assignment is supported by the observation that the predicted 22 N-terminal amino acid residues displays the characteristic features of a signal peptide. The cleavage site for a signal peptidase with the highest probability is located between the glycine and alanine residues, respectively, at position 22 and 23 (Von Heijne, (1983) Eur. J. Biochem. 133, 17-21). The proposed translation initiation codon is preceded by an untranslated region of at least 230 nt.

A continuous vWF cDNA coding sequence of 8,439 bp potentially programs the synthesis of a polypeptide of 2,813 amino acid residues, with a calculated MW of 309 kD. To applicants' knowledge this represents the longest coding sequence determined to date. A computer aided search for (partial) homologous amino acid sequences with other proteins, contained within the NIH Protein Sequence Data Bank, did not reveal any major similarity with other proteins. Furthermore, it has been reported that mature vWF protein is a glycoprotein, containing approximately 15 % carbohydrate residues (Sodetz, J.M. et al., (1979) J. Biol. Chem. 254, 10754-10760). If, it is assumed that the carbohydrate moieties also contribute about 15 % by weight to calculated molecular weight of pro-vWF then the molecular weight of pro-vWF will amount to approximately 350 kD.

Peculiarities of the amino acid sequence of Pre-vWF

A comparison of the predicted amino acid sequence (Figure 3) with the established N-terminal amino acid sequence of mature vWF protein (Hessel, E. et al., (1984) Thrombosis Research 35, 637-651) confirms our earlier assumption (Verweij, C.L. et al., (1985) Nucleic Acids Research 13, 4699-4717, based on Dutch patent application 85.00961) that the vWF precursor protein is considerably larger than the reported 260 kD glycoprotein (Wagner, D.D. et al., (1983) J. Biol. Chem. 258, 2065-2067). Alignment of the predicted amino acid sequence with the N-terminal sequence of the mature (225 kD) vWF protein shows that the nucleotide sequence which codes for mature vWF protein initiates at position 229C. This conclusion implicates that the vWF precursor protein is 763 amino acid residues larger than the mature protein. Obviously, this pro-sequence (calculated MW 81 kD) is removed by protein processing to yield the mature vWF glycoprotein with a molecular weight of about 225 kD.

A homology matrix comparison of the amino acid sequence of the vWF precursor protein reveals a quadruplication of a domain (D1, D2, D3, D4) with a length of about 350 amino acid residues. Part of this domain (D', about 96 amino acids) appears to be present at the N-terminus of mature vWF. Figure 4A shows the alignment of these repetitive domains. A salient feature of the pro-sequence is that it largely consists of a duplication of the D domain (D1, D2). The repeats exhibit a significant conservation of the position of cysteine residues, indicative for a structural similarity of the repeats. Interestingly, the pro-sequence comprises an arginine-glycine-aspartic acid sequence ("RGD" sequence) at position 698 till 702. It has been shown that a tetrapeptide with the indicated amino acid sequence can compete with proteins, harboring a similar sequence, which are involved in cell attachment (Pierschbacher, M.D. et al., (1984) Nature 309, 30-33). It has been noticed that another RGD sequence is present within the C-terminal part of the mature vWF protein (Sadler, J.E. et al., (1985) Proc. Natl. Acad. Sci. USA 82, 6394-6398).

In vitro translation of vWF mRNA

A full length vWF cDNA was assembled in order to demonstrate its coding capacity for a unglycosylated precursor protein with a MW of about 300 kD. Full length vWF cDNA was inserted into plasmid pSP65. This plasmid contains the *S. typhimurium* bacteriophage SP6 promoter which allows in vitro "run off" transcription of cloned DNA sequences, specifically directed by SP6 RNA polymerase (Melton, D.A. et al., (1984) Nucl. Acids Research 12, 7035-7056). Such mRNA preparations can be efficiently translated, using a reticulocyte lysate.

Initially, plasmid pSP6330vWF (Figure 1B) was constructed, harboring a continuous, 6,331 bp vWF cDNA sequence. This plasmid contains the entire coding sequence for mature vWF and in addition a sequence coding for 18 amino acid residues from the C-terminal part of the pro-sequence. Initiation of protein synthesis, directed by RNA transcribed from pSP6330vWF DNA, should occur at the methionine codon 8 amino acids downstream of the N-terminus of mature vWF. Translation of the in vitro synthesized vWF mRNA will then yield an (unglycosylated) polypeptide with a calculated MW of 220 kD. Subsequently, pSP8800vWF (Figure 1B) was constructed harboring the complete coding sequence for pre-vWF. This plasmid will encode a protein with a calculated MW of 309,250 D. The plasmids pSP6330vWF and pSP8800vWF were linearized with, respectively, EcoRI and SalI and transcribed in vitro. The results of the in vitro translation of the various vWF mRNAs are given in Figure 5. The polypeptides encoded by pSP6330vWF DNA display a MW of about 200 kD. The discrepancy of this MW with the calculated MW (220 kD) is probably due to inaccuracy in the MW estimation of large proteins in these gels. The complete coding sequence of pSP8800vWF DNA is translated into a polypeptide with a MW substantially larger than 200 kD. To achieve a more accurate MW estimation for this extraordinarily long polypeptide, we produced partial, overlapping polypeptides derived from selected portions of full length vWF cDNA. To that end pSP8800vWF DNA was digested with BamHI and the transcript (2855 nt long) was translated. It should be noted that 405 nt at the 3' end of this transcript constitute the 5' terminus of the transcript generated from pSP6330vWF cleaved with EcoRI. Hence, an

enumeration of the MWs of the polypeptides, mentioned above, should result in a MW of about 309 kD, after subtracting the common protein region. The protein, derived from the BamHI digested template, has an estimated MW of about 100 kD, while as shown
5 before template pSP6330vWF cleaved with EcoRI yields a product of about 200 kD. Enumeration of these MWs and subtracting the common region (15 kD) results in an estimated MW of 285 kD which is in accord with the calculated MW of 309,250 D. Furthermore, translation of transcripts (1320 nt), derived from pSP8800vWF DNA
10 digested with XhoI, reveals a polypeptide with a MW of 39 kD. This result is in agreement with the assignment of the translation initiation side at position 1 till 4.

From these data applicants conclude that they have constructed a full length vWF cDNA with a coding sequence of 8,439 bp which
15 programs the synthesis of a precursor vWF protein consisting of 2,813 amino acid residues.

DISCUSSION

This invention provides to the construction of a plasmid
20 containing full length vWF cDNA. Nucleotide sequence analysis (this patent application; Verweij, C.L. et al., (1985) Nucleic Acids Research 13, 4699-4717, based on Dutch patent application 85.00961; Sadler, J.E. et al., (1985) Proc. Natl. Acad. Sci. USA 82, 6394-6398) revealed that the length of the assembled vWF cDNA, excluding
25 cDNA derived from the polyA tail, amounts to 8,805 bp. This result is in agreement with the length of vWF mRNA (about 9,000 nt) as determined by Northern blot analysis of endothelial (polyA) RNA (Verweij, C.L. et al., (1985) Nucleic Acids Research 13, 4699-4717, based on Dutch patent application 85.00961). The entire coding
30 sequence for a precursor vWF protein is 8,439 bp, corresponding to an unglycosylated polypeptide with a MW of about 309 kD. The translation initiation site for this protein could be assigned to the ATG codon at position 1 till 4 (see Figure 3). This assignment is in accord with the results of in vitro translation experiments

sequence, which is considerably larger than that of the

MW attributed to the precursor glycoprotein for mature vWF has been reported to display a MW of 260 kD (Wagner, D.D. et al., (1983) Biol. Chem. 258, 2065-2067). The discrepancy with the MW that applicants assign to the precursor protein may be due to inaccuracy of MW estimations by SDS-polyacrylamide gelectrophoresis, inherent to large (glyco)proteins.

The sequence encoding mature vWF initiates at 2,286 bp downstream of the translation initiation codon, as inferred from an alignment of the established N-terminal amino acid sequence of mature vWF (Hessel, B. et al., (1984) Thrombosis Research 35, 637-651) with the predicted amino acid sequence (Figure 3). This 2,286 bp long prepro-sequence was shown to be able to encode a polypeptide with a calculated MW of 83 kD.

Consequently, the pro-vWF protein will be processed by a protease to yield two distinct polypeptides. It is conceivable that the specific cleavage between the arginine and serine residues at positions 763 and 764 occurs within the Weibel-Palade bodies of the endothelial cell, since in the presence of a protease inhibitor (PMSF) a complex can be detected within these organelles of mature vWF with another glycoprotein, designated von Willebrand Antigen II (vW AgII) (Montgomery, R.R. and Zimmerman T.S. 1978 J.Clin.Invest. 61, 1498-1507). Several arguments can be advanced which indicate that it is likely that the pro-sequence of the precursor vWF protein is identical to vW AgII.

i) the MW of the unglycosylated pro-sequence (83 kD) fits with the reported MW of the vW AgII glycoprotein of 98 kD.

ii) it has been demonstrated that both vWF and vW AgII are synthesized by cultured endothelial cells and that these proteins are simultaneously released upon stimulation with 1-desamino-8-D-arginine vasopressin (DDAVP) (McCarroll, D.R. et al, 1984 Blood 63, 532-535).

iii) it has been shown using immunofluorescence techniques that both proteins are localized in the perinuclear region and in the Weibel-Palade bodies (McCarroll, D.R. et al. (1985), J.Clin. Invest. 75, 1089-1095).

iv) both vWF and vW AgII are present in platelets and released together after platelet activation.

v) the levels of vWF and vW AgII protein are linearly associated in plasma and both proteins are deficient in plasma and platelets of a patient with severe von Willebrand's disease.

vi) as mentioned before a complex between vWF and vW AgII can be detected in the presence of a serine protease inhibitor (PMSF) and not in the absence of the inhibitor.

The predicted amino acid sequence of the pro-sequence displays a remarkable structure. It is composed of a duplicated segment of about 350 amino acid residues long. These two segments share 37 % amino acid homology. Furthermore, they exhibit a considerable conservation of similarly located cysteine residues, indicating that structural features have been maintained within these direct repeats. Two copies of this repeat within the pro-sequence are also present within mature vWF, whereas part of this repeat is present at the N-terminus of mature vWF. Internal homologous regions have also been reported by Sadler, J.E. et al, Proc. Natl.Acad.Sci.USA, 82, 6394-6398 (1985), two of which have been duplicated, while one is present in triplicate form (fig. 4B). These repeated sequences span a length of about 1070 amino acid residues within the mature vWF protein. The repeated structures that applicants have found are independent of the ones reported by Sadler, J.E. et al. Proc.Natl. Acad.Sci.USA 82, 6394-6398 (1985). From these data applicants conclude that about 90 % of the precursor vWF protein is constituted of repetitive regions, indicating that the precursor vWF gene has evolved from a series of duplicative events of at least four different regions.

The pro-sequence may participate in the formation of large multimers, composed of vWF dimers (Wagner, D.D. and Marder, V.J. 1984, J. of Cell Biology 99, 2123-2130; Lynch, D.E. et al (1983), The Journal of Biological Chemistry 258, 12757-12760). In this respect it may be relevant to note that pro-vWF is extremely rich in cysteines (8.1 %) which are mainly located in the N- and C-terminal parts of the protein. The cysteine content of the pro-sequence is even higher (8.6 %). It has been shown (Fowler, W.E. et al. J.Clin.

of vWF are required to form rod shape domains, connecting two sub-units. Multimer formation should occur by joining of these dimers, probably again by disulfide bridges at their N-termini to create globular domains. Free sulfhydryl groups of the cysteine residues in the pro-sequence are probably a pre-requisite for the formation of multimers.

The presence of a "RGD(C)" amino acid sequence within the pro-sequence may be indicative for another function of this protein. It has been shown that an RGD containing region on proteins, such as fibronectin and vitronectin carry out a crucial role in the interaction with receptors on a cell surface (Pierschbacher, M.D. and Ruoslahti, E. (1984) *Nature* 309, 30-33); Pytela, R. et al. *Proc. Nat. Acad. Sci. USA* 82, 5766-5770). Those interactions are inhibited by RGD containing peptides. Interaction of mature vWF with activated platelets is also inhibited by RGD containing peptides, suggesting that this region on vWF is involved in platelet binding (Ginsburg, M., 1985 *J. Biol. Chem.* 260, 3931-3936); Haverstick, P.M. et al. 1985, *Blood*). Based on the presence of an RGD sequence both in mature vWF and the pro-sequence, which may be equivalent to vW AgII, and on a striking homology and a structural conservation between these two proteins, applicants assume that the pro-sequence might have a similar function as the mature vWF protein in a specific interaction with particular cell surface receptors.

LEGENDS TO THE FIGURES

Fig. 1 Strategy for the construction of vWF cDNAs, the assembly of full length vWF cDNA and the determination of the nucleotide sequence.

A) vWF mRNA is indicated by a bar; open area, signal peptide coding region; hatched area, pro-sequence coding region; solid area, mature vWF coding region.

The oligonucleotides (20-mers) A (6901-6921), B (4819-4839) and C (2467-2487), which were used for primer-directed cDNA synthesis

and/or as probe for hybridizations, are indicated by small bars. The 575 bp BglIII-BamHI and the 350 bp HindIII-XhoI fragments which were used as probes for colony-screening are indicated by open bars. Below the schematic representation of vWF mRNA the five partial, adjacent vWF cDNAs are given which were used for the assembly of full length vWF cDNA AND FOR NUCLEOTIDE SEQUENCING; The fragments I, II, III, IV and V, which were used in the S1 nuclease protection experiments, are shown above the vWF cDNA insert from which they were derived. The arrows indicate the nucleotide sequencing strategy. In the case of sequence analysis according to the procedure of Maxam and Gilbert (1977), the position of the radioactive labelling is given by a short vertical line at the end of an arrow. The slashes at the end of arrows mean that the end labelling was at a terminus, specified by vector DNA. Only restriction endonuclease sites which are relevant in this study are given. B, BamHI; Bg, BglIII; E, EcoRI; H, HindIII; K, KpnI; M, Mst I; N, NarI; P, PvuII; S, SalI; Sc, SacI; X, XbaI; Xh, XhoI.

B) Assembly of full length vWF cDNA. Plasmid pSP6330vWF contains a 6,331 bp vWF cDNA sequence, extending from the HindIII site (position 2235) till the SacI site (position 8562), subcloned in vector pSP64. Plasmid pSP8800vWF includes full length vWF cDNA, extending from the EcoRI site (see Panel A) till the SacI site (position 8562), subcloned in vector pSP65. Restriction endonuclease sites, delimiting the fragments used for the assembly of full length vWF cDNA, are indicated with an asterix. The EcoRI site at the 5' end of full length vWF cDNA originates from the EcoRI linker, used for the construction of pvWF1330 DNA. The sites for restriction enzymes, which were employed to linearize plasmid DNAs for in vitro "run off" transcription by SP6 RNA polymerase, are indicated by a dot. The SalI site in plasmid pSP8800vWF and the EcoRI site in plasmid pSP6330vWF are present in the polylinkers of the pSP-type vectors.

Fig. 1. S1 nuclease protection analysis of endothelial vWF mRNA.

hybridized with [³²P]-labelled probes, containing vWF cDNA sequences. The construction of the different probes and the conditions used are described in the section Experimental Procedures. The vWF cDNA segments, present in the probes, are shown in Fig. 1.

5 Panel A shows the results after electrophoresis of the samples in a 0.8 % alkaline agarose gel. Panel B gives the results after electrophoresis in a 6 % polyacrylamide - 8 M urea gel. Lanes 1: hybridization with probe III, containing the 1,444 bp vWF cDNA fragment III (Fig. 1). Lanes 2: hybridization with probe V, containing the

10 2,400 bp vWF cDNA fragment V (Fig. 1). Lanes 3: hybridization with probe I which is equivalent to the 585 bp vWF cDNA fragment I (Fig. 1). Lanes 4: hybridization with probe IV, containing the 565 bp vWF cDNA fragment IV (Fig. 1). Lanes 5: hybridization with probe II, containing the 765 bp vWF cDNA fragment II (Fig. 1). Symbols: -,

15 incubation of hybridized components in the absence of S1 nuclease; +, incubation of the hybridized components in the presence of S1 nuclease; c, incubation of the samples with S1 nuclease after hybridization in the absence of endothelial polyA⁺ RNA; M, single stranded DNA length markers.

20

Fig.3 Nucleotide sequence of 8806 bp of vWF cDNA, derived from the 5' terminus of vWF mRNA. The numbering starts at the putative ATG translation initiation codon. The predicted amino acid sequence is shown beneath the nucleotide sequence and are separately numbered,

25 again starting at the putative methionine translation initiation codon. Potential N-linked glycosylation sites are underlined. The tripeptide arginine-glycine-aspartic acid is boxed.

Fig.4 Internal homology within the precursor for vWF.

30 A) Alignment of the amino acid sequences of the four repeated domains D1, D2, D3, D4 and D'. The one-letter notation is used and the amino acids are numbered as indicated in Fig. 3. Residues which are identical among the four or five repeats are boxed.

B) Schematic representation of internal homologous regions within pro-vWF. Indicated are the triplicated domain A (A1, A2 and A3) and

35 two duplicated domains B (B1 and B2) and C (C1 and C2), as reported by Sadler et al. (1985), and the quadruplicated domain D (D1, D2,

D3 and D4). The numeral position of these repeats are listed: A1 (residues 1242 till 1480), A2 (1480-1673), A3 (1673-1875), B1 (2296-2331), B2 (2375-2400), C1 (2400-2516), C2 (2544-2663), D1 (34-387), D2 (387-746), D3 (866-1242), D4 (1947-2299) and D' (769-866).

Fig. 5 In vitro translation of vWF mRNA.

Capped vWF mRNA was prepared in vitro, using "run off" transcription with SP6 RNA polymerase, as described in the section Experimental Procedures. The RNA preparations were added to a reticulocyte lysate translation system, containing [³⁵S]-methionine, and polypeptides were synthesized for 90 min. The polypeptides were fractionated on a 8 % SDS-polyacrylamide gel and then subjected to fluorography. M: MW marker proteins. E: endogeneously synthesized polypeptides (without added RNA). Lane 1: Polypeptides encoded by vWF mRNA transcribed from pSP6330vWF DNA, digested with EcoRI. Lane 2: Polypeptides encoded by vWF mRNA transcribed from pSP8800vWF DNA, digested with SalI. Lane 3: polypeptides encoded by vWF mRNA transcribed from pSP8800vWF DNA, digested with BamHI. Lane 4: polypeptides encoded by vWF mRNA transcribed from pSP8800vWF DNA, digested with XhoI.

A sample of the recombinant DNA plasmid pSP8800vWF in strain E.coli DH 1 was deposited at the "Centraalbureau voor Schimmelcultures" in Baarn, The Netherlands, under number CBS 163.86 on March 26, 1986.

1. cDNA fragment which can be introduced into a recombinant cDNA plasmid or phage, characterized in that the cDNA fragment corresponds at least partially to the gene which codes for the biological activity of the human von Willebrand factor.
2. cDNA fragment according to claim 1, characterized in that the said cDNA fragment has the nucleotide sequence shown in Figure 3 or a part thereof.
3. Recombinant cDNA plasmid or phage, characterized in that the cDNA fragment provided therein corresponds at least partially to the gene which codes for the von Willebrand factor.
4. Recombinant cDNA plasmid or phage according to claim 3, characterized in that the cDNA fragment introduced has the nucleotide sequence shown in Figure 3 or a part thereof.
5. Recombinant cDNA plasmid according to claims 3 or 4, characterized in that the plasmid contains the vector pSP65.
6. Microorganism, animal cell or human cell containing a recombinant cDNA plasmid or phage, characterized in that the recombinant cDNA plasmid or phage is defined in any of the claims 3-5.
7. Microorganism according to claim 6, characterized in that the microorganism is *Escherichia coli*.
8. Microorganism according to claim 7, characterized in that the microorganism is the strain *E. coli* DH 1.
9. Strain *E. coli* DH 1 containing the recombinant cDNA plasmid pSP8800vWF deposited at the Centraalbureau voor Schimmelcultures in Baarn, The Netherlands under C.B.S-number 163.86.
10. Method for the preparation of proteins by the cultivation of a microorganism respectively animal or human cells containing a recombinant cDNA plasmid or phage, characterized by cultivating a host defined in any of the claims 6-9.
11. (Glyco) proteins obtained by the method according to claim 10.
12. vWF (glyco) protein having the amino acid sequence corresponding to the nucleotide sequence of 2518-8667 shown in Figure 3.
13. (Glyco) protein having the amino acid sequence corresponding to the nucleotide sequence of 295-2517 shown in Figure 3.
14. Pharmaceutical composition containing one or more biologically

active (glyco) proteins prepared according to claim 10.

15. Pharmaceutical composition containing the biologically active (glyco) proteins according to claims 12 or 13.

1941

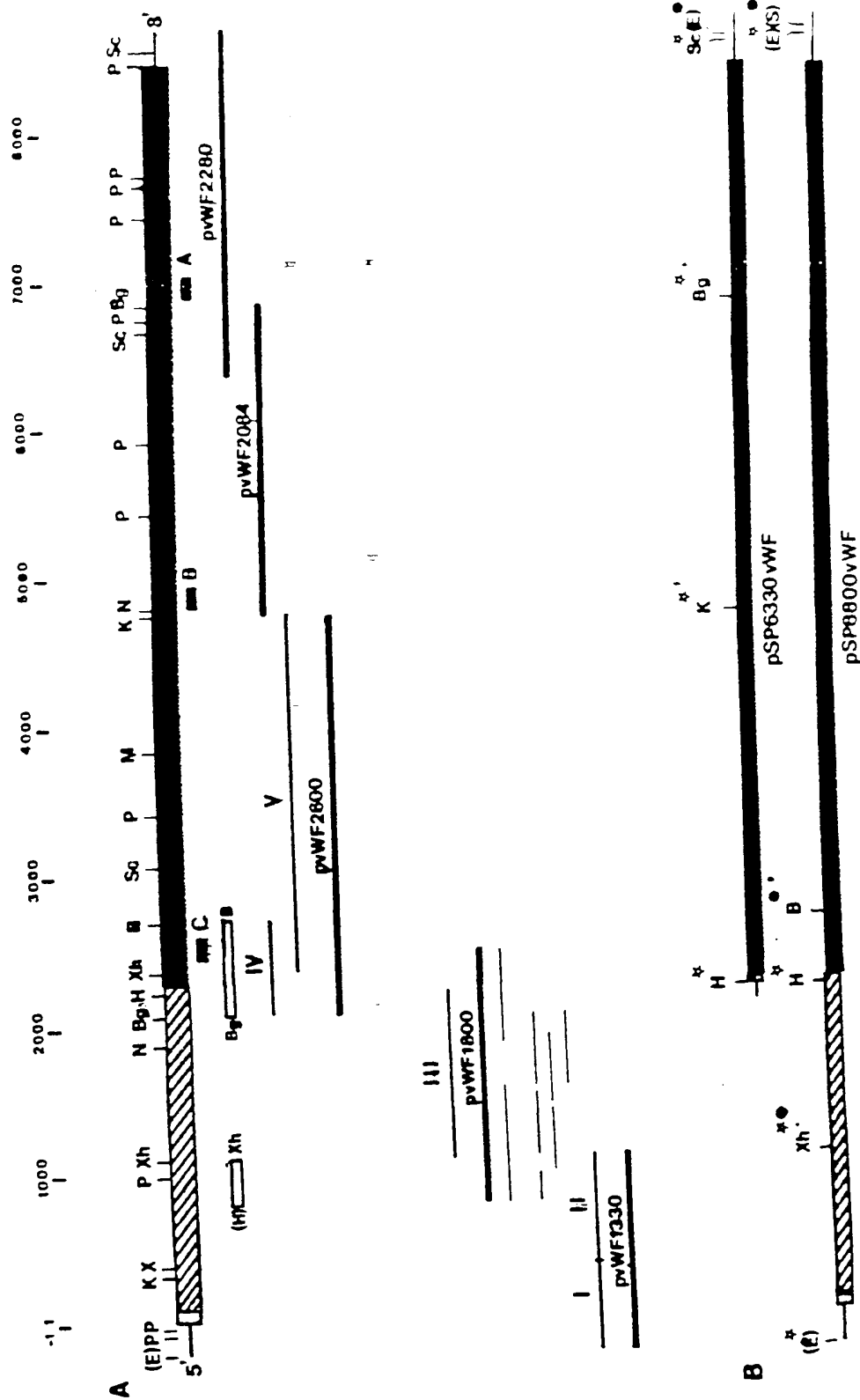


fig-2

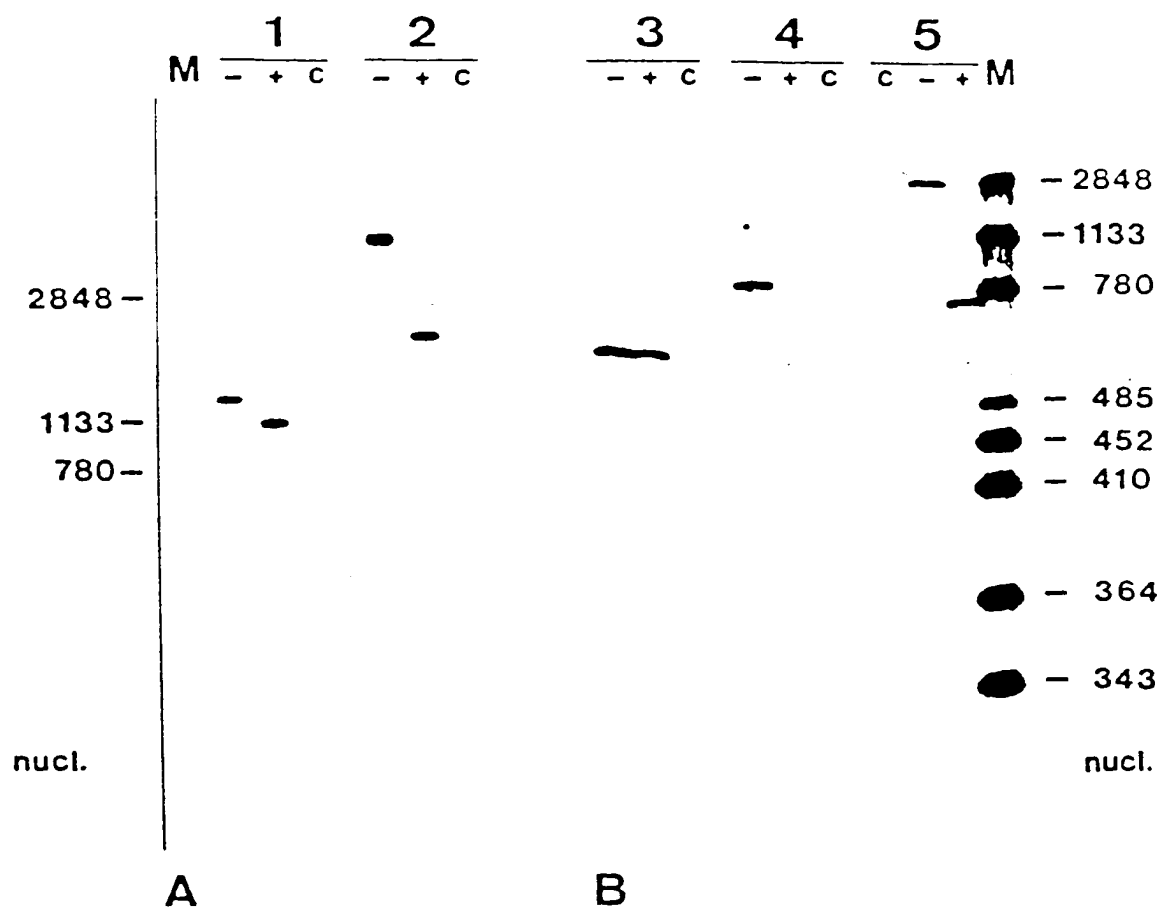


fig - 3⁽¹⁾

0197592

VWF TOTAL

* = M
+ = STOP

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      31                               61
GAAAGGGAGGGTGGTGGTGGATGTCACAGCTTGGGCTTTATCTCCCCAGCAGTGGGAT
      *

      91                               121
TCCACAGCCCCCTGGGCTACATAACAGCAAGACAGTCCGGAGCTGTAGCAGACCTGATTGA
      +

      151                              181
GCCTTTGCAGCAGCTGAGAGCATGGCCTAGGGTGGGCGGCACCATTGTCCAGCAGCTGAG
      * +

      211                              241
TTTCCCAGGGACCTTGGGAGATAGCCGCAGCCCTCATTTGCAGGGGAAGATGATTCTCTGCC
      * I P A

      271                              301
AGATTTCGCGGGGTGCTGCTTGTCTCTGGCCCTCATTTTGCAGGGACCCCTTTGTGCAGAA
R F A G V L L A L A L I L P G T L C A E

      331                              361
GGAAGTCCGCGGCAGGTGATCCACGGCCCCGATGCAGCCCTTTTCGGAAGTGACTTCGTCAAC
G T R G R S S T A R C S L F G S D F V N

      391                              421
ACCTTTGATGGGAGCATGTACAGCTTTGCGGGATACTGCAGTTACCTCCTGGCAGGGGGC
T F D G S * Y S F A G Y C S Y L L A G G

      451                              481
TGCCAGAAACGCTCCTTCTCGATTATTGGGGACTTCCAGAATGGCAAGAGAGTGAGCCTC
C Q K R S F S I I G D F Q N G K R V S L

      511                              541
TCCGTGTATCTTGGGGAATTTTTTGACATCCATTTGTTTGTCAATGGTACCGTGACACAG
S V Y L G E F F D I H L F V N G T V T Q

      571                              601
GGGGACCAAAGAGTCTCCATGCCCTATGCCCTCCAAAGGGCTGTATCTAGAACTGAGGCT
G D Q R V S * P Y A S K G L Y L E T E A

      631                              661
GGGTACTACAAGCTGTCCGGTGAGGCCATGGCTTTGTGGCCAGGATCGATGGCAGCGGC
G Y Y K L S G E A Y G F V A R I D S S G

      691                              721
AACTTTCAAGTCTGCTGTGTCAGACAGATACTTCAACAAGACCTGCGGGCTGTGTGGCAAC
N F Q V L L S D R Y F N K T C G L C G N

      751                              781
TTTAACATCTTTGCTGAAGATGACTTTATGACCCAAGAAGGGACCTTGACCTCGGACCT
F N I F A E D D F * T Q E G T L T S D P

      811                              841
TATGACTTTGCCAACTCATGGGCTCTGAGCAGTGGAGAACAGTGGTGTGAACGGGCATCT
Y D F A N S W A L S S G E Q W C E R A S

      871                              901
CCTCCCAGCAGCTCATGCAACATCTCCTCTGGGGAAATGCAGAAGGGCCTGTGGGAGCAG
P P S S S C N I S S G E * Q K G L W E Q

```


5-1-12

VWF TOTAL

* = M
+ = STOP

931 961
TGCCAGCTTCTGAAGAGCACCTCGGTGTTTGCCCGCTGCCACCCCTCTGGTGGACCCCGAG
C Q L L K S T S V F A R C H P L V D P E

991 1021
CCTTTTGTGGCCCTGTGTGAGAAGACITTTGTGTGAGTGTGCTGGGGGGCTGGAGTGGCGC
P F V A L C E K T L C E C A G G L E C A

1051 1081
TGCCCTGCCCTCCTGGAGTACGCCCGACCTGTGCCAGGAGGGGAATGGTGTGTACGGC
C P A L L E Y A R T C A Q E G * V L Y G

1111 1141
TGGACCGACACAGCGCGTGCAGCCCAAGTGTGCCCTGCTGGTATGGAGTATAGGCAGTGT
W T D H S A C S P V C P A G * E Y R Q C

1171 1201
GTGTCCCTTGCGCCAGGACCTGCCAGACCTGCACATCAATGAAATGTGTGAGGAGCGA
V S P C A R T C Q S L H I N E * C Q E R

1231 1261
TGGCTGGATGGCTGCAGCTGCCCTGAGGGACAGCTCCTGGATGAAGGCCTCTGCGTGGAG
C V D G C S C P E G Q L L D E G L C V E

1291 1321
AGCACCGAGTGTCCCTGCGTGCATTCCGGAAGCGCTACCCCTCCCGGCACCTCCCTCTCT
S T E C P C V H S G K R Y P P G T S L S

1351 1381
CGAGACTGCAACACCTGCATTTGCCGAACAGCCAGTGGATCTGCAGCAATGAAGAATGT
R D C N T C I C R N S Q W I C S N E E C

1411 1441
CCAGGGGAGTGCCTTGTGCAGGTCAATCACACTTCAAGAGCTTTGACAACAGATACTTC
P G E C L V T G Q S H F K S F D N R Y F

1471 1501
ACCTTCAGTGGGATCTGCCAGTACCTGCTGGCCCGGGATTGCCAGGACCACTCCTTCTCC
T F S G I C Q Y L L A R D C Q D H S F S

1531 1561
ATTGTCAATTGAGACTGTCCAGTGTGCTGATGACCGCGACGCTGTGTGCACCCGCTCCGTC
I V I E T V Q C A D D R D A V C T R S V

1591 1621
ACCGTCCGGCTGCCTGGCCTGCACAACAGCCTTGTGAAACTGAAGCATGGGGCAGGAGTT
T V R L P G L H N S L V K L K H G A G V

1651 1681
GCCATGGATGGCCAGGACGTCCAGCTCCCCCTCCTGAAAGGTGACCTCCGCATCCAGCGT
A * D G Q D V Q L P L L K G D L R I Q R

1711 1741
ACAGTGACGGCCTCCGTGCGCCTCAGCTACGGGGAGGACCTGCAGATGGAGTGGGATGGC
T V T A S V R I S Y G F D I O * D W D G

Fig-3 (2)

VWF TOTAL

* = M
+ = STOP

1831 1861
GGGAATTACAATGGCAACCAGSGCGACGACTTCCTTACCCCTCTGGGCTGGCGGAGCCC
G N Y N G N Q G D D F L T P S G L A E P

1891 1921
CGGGTGGAGGACTTCGGGAACGCCTGGAAGCTGCACGGGGACTGCCAGGACCTGCAGAAG
R V E D F G N A W K L H G D C Q D L Q K

1951 1981
CAGCACAGCGATCCCTGCGCCCTCAACCCGCGCATGACCAGGTTCTCCGAGGAGGCGTGC
Q H S D P C A L N P R * T R F S E E A C

2011 2041
GCGGTCTGACGTCCCCACATTGAGGGCTGCCATCGTGCCGTGAGCCCGCTGCCCTAC
A V L T S P T F E A C H R A V S P L P Y

2071 2101
CTGCGGAACCTGCCGCTACGACGTGTGCTCTGCTCGGACGGCCGCGAGTGCTGTGCGGC
L R N C R Y D V C S C S D G R E C L C G

2131 2161
GCCCTGGCCAGCTATGCCGCGGCCTGCGCGGGGAGAGGCGTGCGCGTGCCTGGCGCGAG
A L A S Y A A A C A G R G V R V A W R E

2191 2221
CCAGGCCGCTGTGAGCTGAACCTGCCCCGAAAGGCCAGGTGTACCTGCAGTGCGGGACCCCC
P G R C E L N C P K G Q V Y L Q C G T P

2251 2281
TGCAACCTGACCTGCCGCTCTCTCTTACCCGGATGAGGAATGCAATGAGGCCTGCCTG
C N L T C R S L S Y P D E E C N E A C L

2311 2341
GAGGGCTGCTTCTGCCCCCAGGGCTCTACATGGATGAGAGGGGGGACTGCGTGCCCCAAG
E G C F C P P G L Y * D E R G D C V P K

2371 2401
GCCAGTGCCCTGTTACTATGACGGTGAGATCTTCCAGCCAGAAGACATCTTCTCAGAC
A Q C P C Y Y D G E I F Q P E D I F S D

2431 2461
CATCACACCATGTGCTACTGTGAGGATGGCTTCATGCACTGTACCATGAGTGGAGTCCCC
H H T * C Y C E D G F * H C T * S G V P

2491 2521
GGAAGCTTGCTGCCTGACGCTGTCTCTCAGCAGTCCCCTGTCTCATCGCAGCAAAAGGAGC
G S L L P D A V L S S P L S H R S K R S

2551 2581
CTATCCTGTGCGCCCCCATGGTCAAGCTGGTGTGTCCCGCTGACAACCTGCGGGCTGAA
L S C R P P * V K L V C P A D N L R A E

2611 2641
GGGCTCGAGTGTACCAAAACGTGCCAGAATATGACCTGGAGTGCATGAGCATGGGCTGT
G L E C T K T C Q N Y D L E C * S * G C

2671 2701
GTCTCTGGCTGCCTCTGCCCCCGGGCATGGTCCGGCATGAGAACAGATGTGTGGCCCTG
V S G C L C P P G * V R H E N R C V A L

fig - 3⁽⁴⁾

VWF TOTAL

* = M
- = STOP

```

      2731                               2761
GAAAGGTGTCCCTGCTTCCATCAGGGCAAGGAGTATGCCCTGGAGAAACAGTGAAAGATT
E R C P C F H Q G K E Y A P G E T V K I

      2791                               2821
GGCTGCAACACTTGTGTCTGTGCGGACCGGAAGTGAAGTGCACAGACCATGTGTGTGAT
G C N T C V C R D R K W N C T D H V C D

      2851                               2881
GCCACGTGTCTCCACGATCGGCATGGCCCACTACCTCACCTTCGACGGGCTCAAATACCTG
A T C S T I G * A H Y L T F D G L K Y L

      2911                               2941
TTCCCCGGGGAGTGCCAGTACGTTCTGGTGCAGGATTACTGCGGCAGTAACCCCTGGGACC
F P G E C Q Y V L V Q D Y C G S N P G T

      2971                               3001
TTTCGGATCCTAGTGGGGAATAAGGGATGCAGCCACCCCTCAGTGAAATGCAAGAAACGG
F R I L V G N K G C S H P S V K C K K R

      3031                               3061
GTCACCATCCTGGTGGAGGGAGGAGAGATTGAGCTGTTTGACGGGGAGGTGAATGTGAAG
V T I L V E G G E I E L F D G E V N V K

      3091                               3121
AGGCCCATGAAGGATGAGACTCACTTTGAGGTGGTGGAGTCTGGCCGGTACATCATTCTG
R P * K D E T H F E V V E S G R Y I I L

      3151                               3181
CTGCTGGGCAAGGCCCTCTCCGTGGTCTGGGACCGCCACCTGAGCATCTCCGTGGTCCTG
L L G K A L S V V W D R H L S I S V V L

      3211                               3241
AAGCAGACATACCAGGAGAAAGTGTGTGGCCTGTGTGGGAATTTTGATGGCATCCAGAAC
K Q T Y Q E K V C G L C G N F D G I Q N

      3271                               3301
AATGACCTCACCAGCAGCAACCTCCAAGTGGAGGAGGACCCTGTGGACTTTGGGAAGTCC
N D L T S S N L Q V E E D P V D F G K S

      3331                               3361
TGGGAAGTGAGCTCGCAGTGTGCTGACACCAGAAAAGTGCCTCTGGACTCATCCCTGCC
W E V S S Q C A D T R K V P L D S S P A

      3391                               3421
ACCTGCCATAACAACATCATGAAGCAGACGATGGTGGATTCTCCTGTAGAATCCTTACC
T C H N N I * K Q T * V D S S C R I L T

      3451                               3481
AGTGACGTCTTCCAGGACTGCAACAAGCTGGTGGACCCCGAGCCATATCTGGATGTCTGC
S D V F Q D C N K L V D P E P Y L D V C

      3511                               3541
ATTTACGACACCTGCTCCTGTGAGTCCATTGGGGACTGCGCCTGCTTCTGCGACACCATT
I Y D T C S C E S I G D C A C F C D T I

      3571

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F-5-3(5)

VWF TOTAL

* = M
+ = STDP

3631 3661
TTGTGCCCCAGAGCTGCGAGGAGAGGAATCTCCGCGAGAACGGGTATGAGTGTGAGTGG
L C P Q S C E E R N L R E N G Y E C E W

3691 3721
CGCTATAACAGCTGTGCACCTGCCTGTCAAGTCACGTGTGAGCACCTGAGCCACTGSCC
R Y N S C A P A C Q V T C Q H P E P L A

3751 3781
TGCCCTGTGCAGTGTGTGGAGGGCTGCCATGCCATTGCCCTCCAGGCAAAATCCTGGAT
C P V Q C V E G C H A H C P P G K I L D

3811 3841
GAGCTTTTGCAGACCTGCGTTGACCTGAAGACTGTCCAGTGTGTGAGGTGGCTGSCCG
E L L Q T C V D P E D C P V C E V A G R

3871 3901
CGTTTTGCCCTCAGGAAGAAAGTCACCTTGAATCCAGTGACCTGAGCACTGCCAGATT
R F A S G K K V T L N P S D P E H C Q I

3931 3961
TGCCACTGTGATGTTGTCAACCTCACCTGTGAAGCCTGCCAGGAGCCGGGAGGCCTGGTG
C H C D V V N L T C E A C Q E P G G L V

3991 4021
GTGCCTCCCACAGATGCCCGGTGAGCCCCACCACTCTGTATGTGGAGGACATCTCGSAA
V P P T D A P V S P T T L Y V E D I S E

4051 4081
CCGCCGTTGCACGATTTCTACTGCAGCAGGCTACTGGACCTGGTCTTCCTGCTGGATGGC
P P L H D F Y C S R L L D L V F L L D G

4111 4141
TCCTCCAGGCTGTCCGAGGCTGAGTTTGAAGTGCTGAAGGCCTTTGTGGTGGACATGATG
S S R L S E A E F E V L K A F V V D * *

4171 4201
GAGCGGCTGCGCATCTCCCAGAAGTGGGTCCGCGTGGCCGTGGTGGAGTACCACGACGGC
E R L R I S Q K W V R V A V V E Y H D G

4231 4261
TCCCACGCCCTACATCGGGCTCAAGGACCGGAAGCGACCATCAGAGCTGCGGCGCATTGCC
S H A Y I G L K D R K R P S E L R R I A

4291 4321
AGCCAGGTGAAGTATGCGGGCAGCCAGGTGGCCTCCACCAGCGAGGTCTTGAAATACACA
S Q V K Y A G S Q V A S T S E V L K Y T

4351 4381
CTGTTCCAAATCTTCAGCAAGATCGACCGCCCTGAAGCCTCCCGCATCGCCCTGCTCCTG
L F Q I F S K I D R P E A S R I A L L L

4411 4441
ATGGCCAGCCAGGAGCCCCAACGGATGTCCCGGAACCTTTGTCCGCTACGTCCAGGGCCTG
* A S Q E P Q R * S R N F V R Y V Q G L

4471 4501
AAGAAGAAGAGGTGATTGTGATCCCGGTGGGCATTGGGCCCCATGCCAACCTCAAGCAG
K K K K V I V I P V G I G P H A N L K Q

Fig - 3 (6)

VWF TOTAL

* = M
+ = STOP

4531 4561
 ATCCGCCTCATCGAGAAGCAGGCCCTGAGAACAGSCCTTCGTGCTGAGCAGTGTGGAT
 I R L I E K Q A P E N K A F V L S S V D

4591 4621
 GAGCTGGAGCAGCAAGGGACGAGATCGTTAGCTACCTCTGTGACCTTGCCCTGAGGCC
 E L E Q Q R D E I V S Y L C D L A P E A

4651 4681
 CCTCCTCCTACTCTGCCCGGACATGGCACAAAGTCACTGTGGGCCCGGGGCTCTTGGGG
 P P P T L P P D * A Q V T V G P G L L G

4711 4741
 GTTTCGACCCTGGGGCCCAAGAGGAACCTCCATGGTTCTGGATGTGGCGTTCGTCTGGAA
 V S T L G P K R N S * V L D V A F V L E

4771 4801
 GGATCGGACAAAATTGGTGAAGCCGACTTCAACAGGAGCAAGGAGTTCATGGAGGAGGTG
 G S D K I G E A D F N R S K E F * E E V

4831 4861
 ATTGAGCGGATGGATGTGGGCCAGGACAGCATCCACGTACGGTGTGTCAGTACTCCTAC
 I Q R * D V G Q D S I H V T V L Q Y S Y

4891 4921
 ATGGTGACCGTGGAGTACCCCTTCAGCGAGGCACAGTCCAAAGGGGACATCCTGCAGCGG
 * V T V E Y P F S E A Q S K G D I L Q R

4951 4981
 GTGCGAGAGATCCGCTACCAGGGCGGCAACAGGACCAACACTGGGCTGGCCCTGCGGTAC
 V R E I R Y Q G G N R T N T G L A L R Y

5011 5041
 CTCTCTGACCACAGCTTCTTGGTCAGCCAGGGTGACCGGGAGCAGGCGCCCAACCTG6TC
 L S D H S F L V S Q G D R E Q A P N L V

5071 5101
 TACATGGTCACCGGAAATCCTGCCTCTGATGAGATCAAGAGGCTGCCTGGAGACATCCAG
 Y * V T G N P A S D E I K R L P G D I Q

5131 5161
 GTGGTGCCCATTTGGAGTGGGCCCTAATGCCAACGTGCAGGAGCTGGAGAGGATTGGCTGG
 V V P I G V G P N A N V Q E L E R I G W

5191 5221
 CCCAATGCCCTATCCTCATCCAGGACTTTGAGACGCTCCCCGAGAGGCTCCTGACCTG
 P N A P I L I Q D F E T L P R E A P D L

5251 5281
 GTGCTGCAGAGGCTGCTCCGGAGAGGGGCTGCAGATCCCCACCCTCTCCCCAGCACCT
 V L Q R C C S G E G L Q I P T L S P A P

5311 5341
 GACTGCAGCCAGCCCTGGACGTGATCCTTCTCCTGGATGGCTCCTCCAGTTTCCAGCT
 D C S Q P L D V I L L L D G S S S F P A

5371

Fig - 3 (7)

VWF TOTAL

* = M
+ = STOP

5431 5461
CCTCGTCTCACTCAGGTGTCACTGCTGCAGTATGGAAGCATCACCACCATTGACGTGCCA
P R L T Q V S V L Q Y G S I T T I D V P

5491 5521
TGGACGTGGTCCCGGAGAAAGCCCCATTTGCTGAGCCTTGTGSACGTGCATGCAGCGGGAG
W N V V P E K A H L L S L V D V * Q R E

5551 5581
GGAGGCCCCAGCCAAATCGGGGATGCCTTGGGCTTTGCTGTGCGATACTTGACTTCAGAA
G G P S Q I G D A L G F A V R Y L T S E

5611 5641
ATGCATGGTGCCAGGCCGGGAGCCTCAAAGGCGGTGTCATCCTGGTCACGGACGTCTCT
* H G A R P G A S K A V V I L V T D V S

5671 5701
GTGGATTCACTGGATGCAGCAGCTGATGCCGCCAGGTCCAACAGAGTGACAGTGTCCCT
V D S V D A A A D A A R S N R V T V F P

5731 5761
ATTGGAATTGGAGATCGCTACGATGCAGCCCAGCTACGGATCTTGCCAGGCCCCAGCAGGC
I G I G D R Y D A A Q L R I L A G P A G

5791 5821
GACTCCAACGTGGTGAAGCTCCAGCGAATCGAAGACCTCCCTACCATGGTCACCTTGGGC
D S N V V K L Q R I E D L P T * V T L G

5851 5881
AATTCCTTCTCCACAACTGTGCTCTGGATTTGTTAGGATTTGCATGGATGAGGATGGG
N S F L H K L C S G F V R I C * D E D G

5911 5941
AATGAGAAGAGGCCCGGGGACGTCTGGACCTTGCCAGACCAGTGCCACACCGTGACTTGC
N E K R P G D V W T L P D Q C H T V T C

5971 6001
CAGCCAGATGCCAGACCTTGCTGAAGAGTCATCGGGTCAACTGTGACCGGGGGCTGAGG
Q P D G Q T L L K S H R V N C D R G L R

6031 6061
CCTTCGTGCCCTAACAGCCAGTCCCTGTTAAAGTGAAGAGACCTGTGGCTGCCGCTGG
P S C P N S Q S P V K V E E T C G C R W

6091 6121
ACCTGCCCTGCGTGTGCACAGGCAGCTCCACTCGGCACATCGTGACCTTGTGGGCAG
T C P C V C T G S S T R H I V T F D G Q

6151 6181
AATTTCAAGCTGACTGGCAGCTGTTCTTATGTCCTATTTCAAACAAGGAGCAGGACCTG
N F K L T G S C S Y V L F Q N K E Q D L

6211 6241
GAGGTGATTCTCCATAATGGTGCCTGCAGCCCTGGAGCAAGGCAGGGCTGCATGAAATCC
E V I L H N G A C S P G A R Q G C * K S

6271 6301
ATCGAGGTGAAGCACAGTGCCTCTCCGTGAGCTGCACAGTGACATGGAGGTGACGGTG
I E V K H S A L S V E L H S D * E V T V

Fig - 3 (8)

VWF TOTAL

* = M

+ = STOP

6331 6361
AATGGGAGACTGGTCTCTGTTCTTACGTGGGTGGGAACATGGAAGTCAACGTTTATGGT
N G R L V S V P Y V G G N * E V N V Y G

6391 6421
GCCATCATGCATGAGGTGAGATTCAATCACCTTGGTCACATCTTCACATTCACCTCCACAA
A I * H E V R F N H L G H I F T F T P Q

6451 6481
AACATGAGTTCCAACTGCAGCTCAGCCCCAAGACTTTTGCTTCAAAGACGTATGGTCTG
N N E F Q L Q L S P K T F A S K T Y G L

6511 6541
TGTGGGATCTGTGATGAGAACGGAGCCAATGACTTCATGCTGAGGGATGGCAGTCCACC
C G I C D E N G A N D F * L R D G T V T

6571 6601
ACAGACTGGAAACACTTGTTCAGGAATGGACTGTGCAGCGGCCAGGACAGACGTGCCAG
T D W K T L V Q E W T V Q R P G Q T C Q

6631 6661
CCCATCCTGGAGGAGCAGTGTCTTGTCCCCGACAGCTCCCACTGCCAGGTCTCTCTTTA
P I L E E Q C L V P D S S H C Q V L L L

6691 6721
CCACTGTTTGCTGAATGCCACAAGGTCTGCTCCAGCCACATTCTATGCCATCTGCCAG
P L F A E C H K V L A P A T F Y A I C Q

6751 6781
CAGGACAGTTCGCACCAAGGAGCAAGTGTGTGAGGTGATGCTCTTATGCCACCTCTGT
Q D S S H Q E Q V C E V I A S Y A H L C

6811 6841
CGGACCAACGGGTCTGCTTACTGGAGGACACCTGATTTCTGTGCTATGTCATGCCCA
R T N G V C V D W R T P D F C A * S C P

6871 6901
CCATCTCTGGTCTACAACCACTGTGAGCATGGCTGTCCCCGGCACTGTGATGGCAACGTG
P S L V Y N H C E H G C P R H C D G N V

6931 6961
AGCTCCTGTGGGGACCATCCCTCCGAAGGCTGTTTCTGCCCTCCAGATAAAGTCATGTTG
S S C G D H P S E G C F C P P D K V * L

6991 7021
GAAGGCAGCTGTGTCCCTGAAGAGGCTGCACTCAGTGCATTGGTGAGGATGGAGTCCAG
E G S C V P E E A C T Q C I G E D G V Q

7051 7081
CACCAGTTCCTGGAAGCCTGGGTCCCGGACCACCGCCCTGTCAGATCTGCACATGCCTG
H Q F L E A W V P D H Q P C Q I C T C L

7111 7141
AGCGGGCGGAAGGTCAACTGCACAACGCAGCCCTGCCCCACGGCCAAAGCTCCACGTGT
S G R K V N C T T Q P C P T A K A P T C

7171

fig- 3⁽⁹⁾

VWF TOTAL

* = M

+ = STOP

6331 6361
 AATGGGAGACTGGTCTCTGTTTCCTTACGTGGGTGGGAACATGGAAGTCAACGTTTATGGT
 N G R L V S V P Y V G G N * E V N V Y G

6391 6421
 GCCATCATGCATGAGGTGAGATTCAATCACCTTGGTCACATCTTCACATTCACCTCCACAA
 A I * H E V R F N H L G H I F T F T P Q

6451 6481
 AACAAATGAGTTCCAACTGCAGCTCAGCCCCAAGACTTTTGCTTCAAAGACGTATGGTCTG
 N N E F Q L Q L S P K T F A S K T Y G L

6511 6541
 TGTGGGATCTGTGATGAGAACGGAGCCAAATGACTTCATGCTGAGGGATGGCAGATCCAG
 C G I C D E N G A N D F * L R D G T V T

6571 6601
 ACAGACTGGAAAACACTTGTTCAGGAATGGACTGTGCAGCGGCCAGGACAGCGTGCCAG
 T D W K T L V Q E W T V Q R P G Q T C Q

6631 6661
 CCCATCCTGGAGGAGCAGTGTCTTGTCCCCGACAGCTCCCACTGCCAGGTCTCTCTTA
 P I L E E Q C L V P D S S H C Q V L L L

6691 6721
 CCACTGTTTGCTGAATGCCACAAGGTCTGGCTCCAGCCACATTCTATGCCATCTGCCAG
 P L F A E C H K V L A P A T F Y A I C Q

6751 6781
 CAGGACAGTTGCGACCAAGGAGCAAGTGTGTGAGGTGATCGCCTCTTATGCCACCTCTGT
 Q D S S H Q E Q V C E V I A S Y A H L C

6811 6841
 CGGACCAACGGGTCTGCGTTGACTGGAGGACACCTGATTTCTGTGCTATGTCATGCCCA
 R T N G V C V D W R T P D F C A * S C P

6871 6901
 CCATCTCTGGTCTACAACCACTGTGAGCATGGCTGTCCCCGGCACTGTGATGGCAACGTG
 P S L V Y N H C E H G C P R H C D G N V

6931 6961
 AGCTCCTGTGGGGACCATCCCTCCGAAGGCTGTTTCTGCCCTCCAGATAAAGTCATGTTG
 S S C G D H P S E G C F C P P D K V * L

6991 7021
 GAAGGCAGCTGTGTCCCTGAAGAGGCCTGCACTCAGTGCATTGGTGAGGATGGAGTCCAG
 E G S C V P E E A C T Q C I G E D G V Q

7051 7081
 CACCACTTCTGGGAAGCCTGGGTCCCGGACCAAGCCCTGTCAGATCTGCACATGCCTG
 H Q F L E A W V P D H Q P C Q I C T C L

7111 7141
 AGCGGGCGGAAGGTCAACTGCACAACGCCCTGCCCAAGGCCAAAGTCCCACGTGT
 S G R K V N C T T Q P C P T A K A P T C

7171 7201
 GGCCTGTGTGAAGTAGCCCCGCTCCGCGAGAATGCAGACCAAGTGTGCCCCGAGTATGAG
 G L C E V A R L R Q N A D Q C C P E Y E

75-3 (13)

VWF TOTAL

* = M
+ = STOP

8131 8161
AACCCCTGCCCCCTGGGTTACAAGGAAGAAAATAACACAGGTGAATGTTGTGGGAGATGT
N P C P L G Y K E E N N T G E C C G R C

8191 8221
TTGCCTACGGCTTGACCATTCAGCTAAGAGGAGGACAGATCATGACACTGAAGCGTGAT
L P T A C T I Q L R G G Q I * T L K R D

8251 8281
GAGACGCTCCAGGATGGCTGTGATACTCACTTCTGCAAGGTCAATGAGAGAGGAGAGTAC
E T L Q D G C D T H F C K V N E R G E Y

8311 8341
TTCTGGGAGAAGAGGGTCACAGGCTGCCACCCTTTGATGAACACAAGTGTCTGGCTGAG
F W E K R V T G C P P F D E H K C L A E

8371 8401
GGAGGTAAATATGAAATTCACAGGCACCTGCTGTGACACATGTGAGGAL TGAGTCC
G G K I * K I P G T C C D T C E E P E S

8431 8461
AACGACATCACTGCCAGGCTGCAGTATGTCAAGGTGGGAAGCTGTAAGTCTGAAGTAGAG
N D I T A R L Q Y V K V G S C K S E V E

8491 8521
GTGGATATCCACTACTGCCAGGGCAAATGTGCCAGCAAAGCCATGTACTCCATTGACATC
V D I H Y C Q G K C A S K A * Y S I D I

8551 8581
AACGATGTGCAGGACCAAGTGCTCCTGTGCTCTCCGACACGGACGGAGCCCATGCAGGTG
N D V Q D Q C S C C S P T R T E P * Q V

8611 8641
GCCCTGCACCTGCACCAATGGCTCTGTTGTGTACCATCAGGTTCTCAATGCCATGGAGTGC
A L H C T N G S V V Y H Q V L N A * E C

8671 8701
AAATGCTCCCCAGGAAGTCCAGCAAGTGAGGCTGCTGCAGCTGCATGGGTGCCTGCTGC
K C S P R K S S K +

8731 8761
TGCTGCCTTGCCCTGATGGCCAGGCCAGAGTGCTGCCAGTCTCTGCATGTTCTGCTCT
*

8791
TGTGCCCTTCTGAGCCCAATAAAGGCTGAGCTCTTATCTTGCA
+

fig-4L

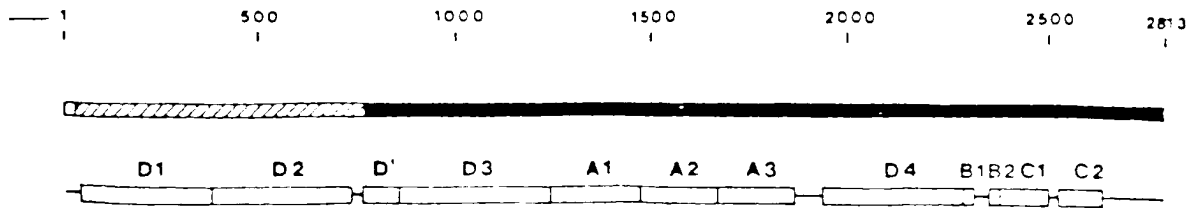
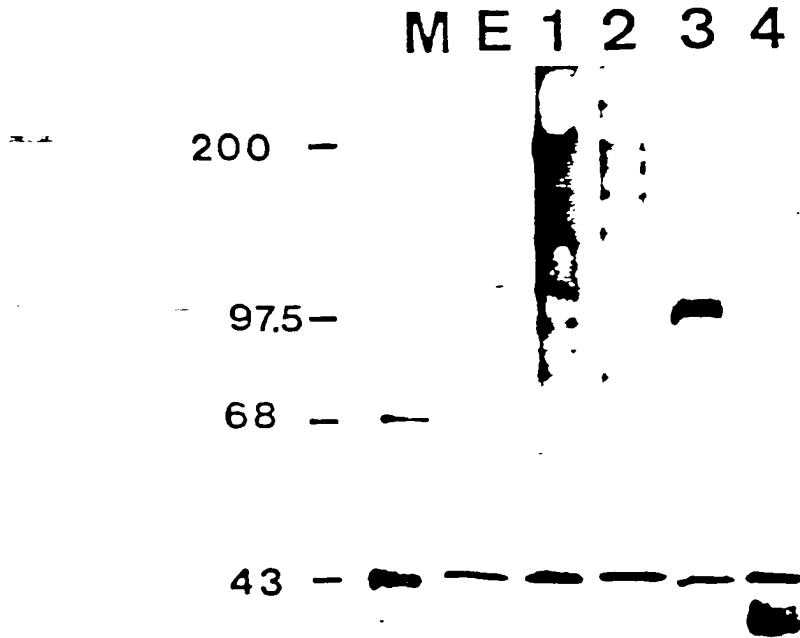


fig-5





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Place of search THE HAGUE		Date of completion of the search 09-07-1986	Examiner DELANGHE L. L. M.
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